

Immunotoxicological Effects of Agent Orange Exposure to the Vietnam War Korean Veterans

Hyoung-Ah KIM¹, Eun-Mi KIM², Yeong-Chul PARK², Ji-Yeon YU³, Seung-Kwon HONG³,
Seong-Hoon JEON⁴, Kui-Lea PARK⁵, Sook-Jin HUR⁶ and Yong HEO^{2*}

¹Department of Preventive Medicine, College of Medicine, The Catholic University of Korea, 505 Banpo-dong, Seoul, Korea

²Department of Occupational Health, College of Natural Sciences, Catholic University of Daegu, 330 Kumrak 1-ri, Hayang-eup, Kyongsan-si, Kyongbuk, Korea

³School of Public Health, Seoul National University, 28 Yeonkun-dong, Seoul, Korea

⁴College of Medicine Asan Medical Center, University of Ulsan, 388-1 Poongnap-dong, Seoul, Korea

⁵National Institute of Toxicological Research, Korea Food & Drug Administration, 5 Nokbun-dong, Seoul, Korea

⁶Division of Bacterial Products, Korea Food & Drug Administration, 5 Nokbun-dong, Seoul, Korea

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Abstract: Immunomodulatory effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) demonstrated using animals are thymic atrophy, downregulation of cytotoxic T or B lymphocyte differentiation or activation, whereas human immunotoxicities have not been investigated well. This study was undertaken to evaluate overall immunologic spectrum of the Vietnam War Korean veterans exposed to Agent Orange contaminated with TCDD. Quantity of red blood cells, hemoglobin and hematocrit in the veterans suffered from chronic diseases associated with Agent Orange exposure (Veterans-patient group) were decreased in comparison with those of the veterans without the diseases and the age-matched healthy controls, but no differences in leukocyte populations. Plasma IgG levels were lowered in the veterans than the controls, owing to significant decrease in the IgG1 levels. Increase in the IgE levels was observed in the plasma from the veterans. Alteration of T cell-mediated immunity was also resulted from activation of peripheral blood mononuclear cells with polyclonal T cell activators. Production of IFN γ , a major cytokine mediating host resistance against infection or tumoregenesis, was lowered in the veterans-patient group. However, production of IL-4 and IL-10, representative cytokines involved with hypersensitivity induction, was enhanced in the patient group. Overall, this study suggests that military service in Vietnam and/or Agent Orange exposure disturbs immune-homeostasis resulting in dysregulation of B and T cell activities.

Key words: Agent Orange, Vietnam War Veterans, RBC, Immunoglobulins, Cytokines, Autoantibodies

Introduction

The Vietnam War Korean veterans, who did military services from 1964 to 1973 in Vietnam, could be exposed to Agent Orange contaminated with TCDD through participation on spraying Agent Orange. Amount of Agent Orange sprayed by Korean soldiers is estimated as much as

500 thousand gallons¹). Among approximately 320 thousand Korean soldiers dispatched to Vietnam, over 90 thousand veterans have reported to have health problems related with Agent Orange exposure. Number of the veterans registered as the patient definitely connected with Agent Orange exposure is currently 17,711 veterans, and 46,249 veterans as the patient probably associated with the exposure²). Based on the results of two Governmental epidemiologic studies, peripheral nerve disease is the most prevalent disease

*To whom correspondence should be addressed.

followed by lung cancer, Buerger's disease, larynx cancer, non-Hodgkin's lymphoma, and chloracne in the veterans registered as the patient connected with Agent Orange exposure^{1,3}. Concerning on the veterans enrolled as the patient probably associated with Agent Orange exposure, hypertension was the most prevalent disease followed by diabetes mellitus, seborrheic dermatitis, central nervous diseases, liver diseases, multifocal neuropathy, cancer, hyperlipidemia, cerebrovascular disease, ischemic heart disease, and other skin diseases such as chronic urticaria, psoriasis vulgaris. Considering pathogenesis involved with the diseases addressed above, immune system is assumed to be a potential target of TCDD contained in the Agent Orange, but no systemic investigation was performed to evaluate magnitude of immunotoxicities in the Korean veterans exposed to Agent Orange.

TCDD has been reported to suppress humoral immunity associated with inhibition of B cell differentiation and activation in experimental animals⁴⁻⁶. Furthermore, immunotoxicities on rodent T cells have been demonstrated, i.e., suppression of cytotoxic T lymphocyte activity, enhancement of activation-induced T cell death, or inhibition of helper T cell-derived cytokines⁷⁻⁹. However, human studies did not show a consistent relationship between TCDD exposure and alteration in immune response^{10,11}. This inconsistency may be derived from differences in study details, e.g., characteristics on study subjects, magnitude of TCDD exposure, interval from exposure to study, or criteria for control selection, and so forth. Therefore, we investigated for the first time in Korea whether exposure to Agent Orange modifies immunological spectrum of the Vietnam War veterans. The results obtained show that military service in Vietnam and/or Agent Orange exposure may contribute toward alteration in distribution of plasma IgG subclasses, T cell subset activity, and qualitative characteristics of red blood cells (RBC).

Materials and Methods

Subjects

Study subjects were classified into three groups based on the Agent Orange exposure history and their health status. The **veterans-patient** group was composed of veterans suffering from chronic illness and exposed to 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) during their operation in various areas of Vietnam such as Nha Trang, Cam Ranh, Qui Nhon, Tuy Hoa, Nin Hoa, or Da Nang where Agent Orange contaminated with TCDD had been sprayed during the Vietnam War. They were randomly selected among the

Vietnam veterans who visited a veterans hospital in Seoul for the Agent Orange Registry health examination. The **veterans-normal** group was composed of veterans, who had military service in the various locations described above during the war, but no chronic illness associated with Agent Orange exposure. **Control** volunteers, age-matched (± 5 years) with the veterans-patient subjects, were chosen among healthy subjects visiting a general hospital in Seoul for routine health examination and with no Vietnam War military service. Number of study subjects for each group was twenty-four, twenty-seven, and thirty-six, and their age (mean \pm SD) was 55.3 ± 4.3 , 55.2 ± 3.43 , and 48.2 ± 2.9 years for the veterans-patient, the veterans-normal, and the control, respectively. Informed consent was obtained from each subject prior to their participation in the study. Health status of the participants were investigated clinically by specialist of rehabilitation medicine or family medicine. All sampling procedures complied with the institutional guidelines governing the use of human subjects in research by the Institute for Occupational and Environmental Health, Korea where the correspondence author was employed at the beginning of this study.

Peripheral blood analysis

Hematological parameters including number and proportion of white blood cell (WBC), red blood cells (RBC), lymphocyte, monocyte, granulocyte, platelet were determined using Coulter counter (Beckman coulter, Fullerton, CA). Hemoglobin concentrations and hematocrit values were also measured.

For investigation on lymphocyte activities, peripheral blood mononuclear cells (PBMC) were isolated by Ficoll-Hypaque density gradient centrifugation (Amersham Pharmacia Biotech, Upsala, Sweden). PBMC (1×10^6) were stimulated with 5 ng phorbol 12-myristate 13-acetate (PMA, Sigma, Saint Louis, MO) plus 500 ng ionomycin (Sigma), or 5 μ g phytohemagglutinin (PHA, Sigma) alone for 48 or 72 hours. The highest levels of cytokine production were observed at the stated concentrations of stimulants, and culture supernatants collected at 72 h were considered more reproducible and more representative owing to higher production of IL-4 and IFN γ (data not shown). Culture medium was RPMI 1640 supplemented with nonessential amino acids (1 mM), sodium pyruvate (1 mM), sodium bicarbonate (1%) from Biowhittaker (Walkersville, MD), glutamine (2 mM), 50 μ M 2-mercaptoethanol from Sigma, 1% penicillin-streptomycin-fungizone mixture (Gibco-BRL, Grand Island, NY), and heat-inactivated fetal bovine serum (10%, FBS, Gibco-BRL).

Determination of cytokine concentration

Interleukin-4 (IL-4), IL-10, IFN γ , and Tumor necrosis factor (TNF) α in culture supernatants were assayed by a sandwich ELISA using the method supplied by PharMingen (San Diego, CA) and previously described¹². The capture and detection mAbs pairs were as follows: IL-4, 8D4-8/MP4-25D2; IL-10, JES3-19F1/JES3-12G8; IFN γ , NIB42/4S.B3; TNF α , MAb1/MAb11. The plates (Immulon IV, Dynatech, Chantilly, VA) were read using an ELISA reader (Bio-Tek EL800, Winooski, VT) at 405 nm, which automatically calculated the concentration of cytokines from the standard curves. The lower limit of detection was 15 pg/ml for IL-4, 30 pg/ml for IL-10, 100 pg/ml for IFN γ , and 60 pg/ml for TNF α .

Quantification of plasma IgG subclass

The level of IgG subclasses in the plasma was measured by subclass-specific ELISA as previously described with minor modification^{13, 14}. Briefly, Immulon II plates (Dynatech) were coated with capture antibody (0.2 μ g/100 μ l/well) by overnight incubation at 4°C. Then the plates were blocked with PBS containing 1% bovine serum albumin (BSA) for 2 h at room temperature. Plasma was added to duplicate wells (100 μ l/well) at 1:10000 for IgG1, 1:50000 for IgG2, 1:5000 for IgG3, and 1:25000 dilution for IgG4 with 1% BSA-PBS followed by overnight incubation at 4°C. The bound IgG subclass was detected by adding biotin-mouse anti-human IgG (0.1 μ g/100 μ l/well, PharMingen) for 2 h at room temperature. The plates were washed and further incubated with avidin-peroxidase (0.25 μ g/100 μ l/well, Sigma) for 1 h. Finally, bound enzyme activity was measured using the substrate 2,2'-azino-bis (3-ethylbenzthiazolone-6-sulfonic acid, Sigma). The optical density was read at 405 nm. The sensitivity of the assays was 5 ng/ml for IgG1, 15 ng/ml for IgG2 and IgG3, and 1.5 ng/ml for IgG4. Quality control for the assays was performed using WHO reference serum (National Institute for Biological Standards and Control, Hertfordshire, UK) and Human IgG subclass profile ELISA kit (Zymed Laboratories, South San Francisco, CA).

Evaluation of plasma IgE and autoantibody levels

Quantification of IgE was performed using ELISA kit (IBL Immuno-Biological Laboratories, Hamburg, Germany). Levels of plasma anti-double stranded DNA (ds DNA) was determined using ELISA kit supplied by Scimedx corporation (Denville, NJ) and antibody against the extractable nuclear antigens (ENA) with ELISA kit by BL Diagnostika (Mainz, Germany). The plates were read at 450 nm.

Statistical analysis

Data were initially evaluated for nominal distribution. Statistical significances among groups were tested using Sigmaplot (SPSS Inc., Chicago, IL, USA) by single-factor ANOVA and Dunnett's t-test or Kruskal-Wallis ANOVA and Dunn's test, depending on normality of data. The significances were further confirmed by Student's t-test or Mann-Whitney test. Differences were considered significant when P was less than 0.05. Logarithmic transformation was used for statistical analyses on plasma concentrations of IgG subclass.

Results

Description on diseases diagnosed to the veterans

The veterans-patient group is composed of the Vietnam War veterans diagnosed to have chronic diseases associated with Agent Orange exposure. Among them, twelve veterans (50%) had cardiovascular diseases such as hypertension, hemiplegia, cerebral infarction, telangiectasis, or total hip replacement due to avascular necrosis, and five veterans (21%) were suffered from nervous system disorders such as lumbar radiculopathy, peripheral neuropathy, or cervical myelopathy (data not shown). Dermal pathogenesis including folliculitis, seborrheic dermatitis, or eczema nummulare were also evident to four veterans (17%). In addition, diseases at endocrine, musculo-skeletal, neurologic, or respiratory system were diagnosed to eight veterans. Spinal cord tumor or lipoma was present in one subject, respectively. The patient subjects had 1.5 chronic diseases per person.

Monitoring of hematological alterations

Concerning composition and quantity of peripheral blood cells, white blood cells from the veterans-patient group were not much different from those of other groups. Average number of WBC was $6.81 \pm 0.26 \times 10^3/\mu$ l for the veterans-patient group, $6.74 \pm 0.30 \times 10^3/\mu$ l for the veterans-normal group, and $6.41 \pm 0.22 \times 10^3/\mu$ l for the control group. Number of lymphocyte ($2.32 \pm 0.11 \times 10^3/\mu$ l), monocyte ($0.53 \pm 0.03 \times 10^3/\mu$ l), or granulocyte ($3.96 \pm 0.17 \times 10^3/\mu$ l) for the veterans-patient group was not significantly different from those of other groups. Furthermore, we could not find any significant difference in proportion of lymphocyte, monocyte, and granulocyte within leukocyte population (data not shown).

We also compared parameters related with RBC. Average number of RBC ($4.42 \pm 0.10 \times 10^6/\mu$ l) for the veterans-patient group was significantly lower than those of other groups (Table 1). Even though hemoglobin concentrations and hematocrit values for the veterans-patient subjects were

Table 1. Lowered RBC parameters in the veterans in comparison with those of the control group

Group	RBC parameters (mean ± SD)		
	Number of RBC (10 ⁶ cells/ μ l)	Hemoglobin (g/dl)	Hematocrit (%)
Veterans-patient	4.42 ± 0.10*	14.28 ± 0.35*	41.48 ± 1.00*
Veterans-normal	4.67 ± 0.08	14.79 ± 0.29	43.33 ± 0.76
Control	4.68 ± 0.06	15.32 ± 0.17	44.39 ± 0.51

Statistically significant differences (*) from the veterans-normal and the control group.

Table 2. Alterations in the levels (mean ± SD) of plasma IgE and IgG1

Group	IgE (ng/ml)	IgG subclasses (mg/ml)			
		IgG1	IgG2	IgG3	IgG4
Veterans-patient	1467 ± 351*	4.11 ± 0.74**	4.47 ± 0.44	0.81 ± 0.07	0.22 ± 0.04
Veterans-normal	1693 ± 577*	5.76 ± 0.92	4.06 ± 0.32	0.94 ± 0.09	0.33 ± 0.06
Control	746 ± 178	8.75 ± 1.18	5.73 ± 0.64	0.88 ± 0.09	0.33 ± 0.04

Statistically significant differences (*) from the control group and significantly lowered IgG1 level (**) in the veterans-patient group compared with that of the other two groups.

within the published normal range (12–18 g/dl for hemoglobin and 36–54% for hematocrit)¹⁵, level of these parameters was significantly lower in the veterans-patient group than in the other two groups.

Alteration in humoral immunity following Agent Orange exposure

To monitor potential adverse effects on the humoral immunity due to Agent Orange exposure, we determined the plasma immunoglobulin (IgG subclasses and IgE) levels of the veterans-patient subjects and compared them with those of the veterans-normal and the control subjects. IgG is a mediator of humoral immunity against bacterial or viral infection, and normal levels usually reflect normal function of B cells. Abnormal levels of serum IgE and IgG subclass have been reported in various diseases including allergic diseases such as asthma or atopic dermatitis^{16,17}. The plasma IgE level was significantly enhanced in the veterans-patient subjects compared with that of the control subjects (Table 2). However the significantly higher plasma IgE level of the veterans-patient group may not be associated with the diseases occurred in the patients, in that no significant difference was found between the veterans-patient and the veterans-normal group.

Total IgG levels were significantly different among the groups (the veterans-patient, 9.9 ± 1.0; the veterans-normal, 11.1 ± 1.1; the control, 16.0 ± 1.5 mg/ml). Plasma IgG1

level may be responsible for the difference in the total IgG levels since IgG1 level was significantly lower in the veterans-patient group (4.1 ± 0.7 mg/ml) than in the veterans-normal (5.8 ± 0.9 mg/ml) and the control (8.8 ± 1.2 mg/ml) group (Table 2), whereas no significant alteration was demonstrated with levels of IgG2, IgG3, and IgG4 among the groups.

Alteration of T cell-mediated immunity in the veterans exposed to Agent Orange

Exposure to environmental toxicants has been reported to disturb the balance between the regulatory and effector T cells, which can lead to selective activation or suppression of T cell subsets with production of distinct cytokines resulting in various immunopathologic states in human^{5, 18, 19}. To determine whether Agent Orange exposure could direct alteration in the functional activities of T lymphocytes, levels of cytokines produced *in vitro* from T cells obtained from the veterans-patient subjects were evaluated and compared them with those of the veterans-normal and the control subjects. Higher production of IL-4 and TNF α was achieved through stimulation with PMA plus ionomycin than with PHA, whereas production of IL-10 and IFN γ was higher with PHA stimulation than PMA plus ionomycin. Hereafter we present only the data on IL-4 and TNF α production from T lymphocytes activated with PMA plus ionomycin, vice versa the data on IL-10 and IFN γ production from PHA-

Table 3. Downregulatory production of IFN γ and moderately enhanced production of IL-4 and IL-10 from PBMC stimulated with polyclonal T cell activators in the veterans-patient subjects

Group	Cytokine concentration (pg/ml)				
	IFN γ	TNF α	IL-4	IL-10	IL-4: IFN γ Ratio
Veterans-patient	11164 \pm 4103*	7027 \pm 1940	151 \pm 78**	2421 \pm 393	4.95 \pm 1.72**
Veterans-normal	21113 \pm 1599	10049 \pm 1123	125 \pm 23**	2156 \pm 116	7.04 \pm 1.62**
Control	30378 \pm 8475	6145 \pm 2961	57 \pm 20	1898 \pm 68	1.43 \pm 0.68

PBMC were stimulated with PHA or PMA plus ionomycin for 3 days. The results are expressed as the means \pm SD of cytokines in the culture supernatants. The ratio (mean \pm SD) was calculated by dividing the amount of IL-4 by the amount of IFN γ production in the same subjects followed by multiplication of 1000. Statistically significant difference (*) from the veterans-normal and the control group, and significant differences (**) from the control.

activated T lymphocytes.

IFN γ production was significantly lower in the veterans-patient group than the other groups (Table 3). Even though IFN γ level in the veterans-normal group was higher than that in the veterans-patient group, the IFN γ production remained lower than the control group. This result implies that exposure to Agent Orange plays a downregulatory role for IFN γ production. With regard to TNF α production, no significant difference was observed among the groups. We also investigated the effect of Agent Orange exposure on the production of IL-4 and IL-10, which are known as type-2 cytokines produced from type-2 T cells (Th2 and Tc2)²⁰. IL-4 production was elevated in the veterans exposed to Agent Orange in comparison with that of the control subjects (Table 3), but no difference between the veterans-patient and the veterans-normal group was found. Like-wise, production of IL-10 was upregulated in the veterans in comparison with that of the control subjects without statistical significances. To examine the relative increase in IL-4 to IFN γ production by PBMC T cells from the Agent Orange-exposed individual subjects, the IL-4: IFN γ ratio was calculated (Table 3). The mean ratio for the Agent Orange-exposed was significantly higher (veterans-patient, 4.95; veterans-normal, 7.04) than for the control subjects (1.43).

Quantitative determination of autoantibodies

In order to monitor potential induction of autoimmunity such as Systemic Lupus Erythematosus, or systemic rheumatic diseases in the veterans exposed to Agent Orange^{21, 22}, we evaluated plasma levels of antibody against ds DNA or extractable nuclear antigens. According to the manufacturer's guideline, titer of anti-ds DNA over 25 International Unit (IU) or anti-ENA over 1 Index Value (IV) is considered positive, and titer over 60 IU or 1.2 IV is considered definitely positive. Mean value of the anti-ds DNA antibodies was 17.3 \pm 5.7 and 21.5 \pm 10.1 IU in the veterans-patient and

the veterans-normal subjects, respectively, without significant difference (Fig. 1, left). Mean level of anti-ENA antibodies was not much different between the two groups (0.21 \pm 0.38 for the veterans-patient, 0.15 \pm 0.11 IV for the veterans-normal group, Fig. 1, right). One subject strong positive to anti-ENA antibody was belonging to the veterans-patient group and suffering from total hip replacement due to avascular necrosis. However, the high titer (2.0) of anti-ENA antibody may not be associated with the disease since the other two veterans with the same disease demonstrated titers below 0.25 IV. The data indicates that autoimmune diseases involved with anti-ds DNA or anti-ENA antibody are not resulted from Agent Orange exposure.

Discussion

Toxicities related with TCDD exposure had been introduced to us through investigation on health outcomes following a herbicide factory explosion at Seveso, Italy in 1976²³. According to a recent report on the Seveso accident¹⁰, plasma IgG levels were decreased with increasing TCDD concentration in the plasma, which were measured about 20 years after the accident. Our present study also demonstrated that plasma total IgG levels were significantly decreased in the veterans exposed to Agent Orange (Table 2). In addition, we found that the decrease in IgG levels were attributable to decrease in the levels of IgG1 subclass. Since IgG1 plays a major role for humoral immune response through complement fixation, neutralization, or Fc receptor-mediated phagocytosis, the IgG1 decrease may lead to downregulation of antibody-mediated host resistance, which could enhance an opportunity of infection or tumor progression¹⁶. This finding is well consistent with a German study reporting a significant decrease in the plasma concentration of IgG1 in workers exposed to TCDD²⁴. Furthermore, the levels of plasma IgE, a representative

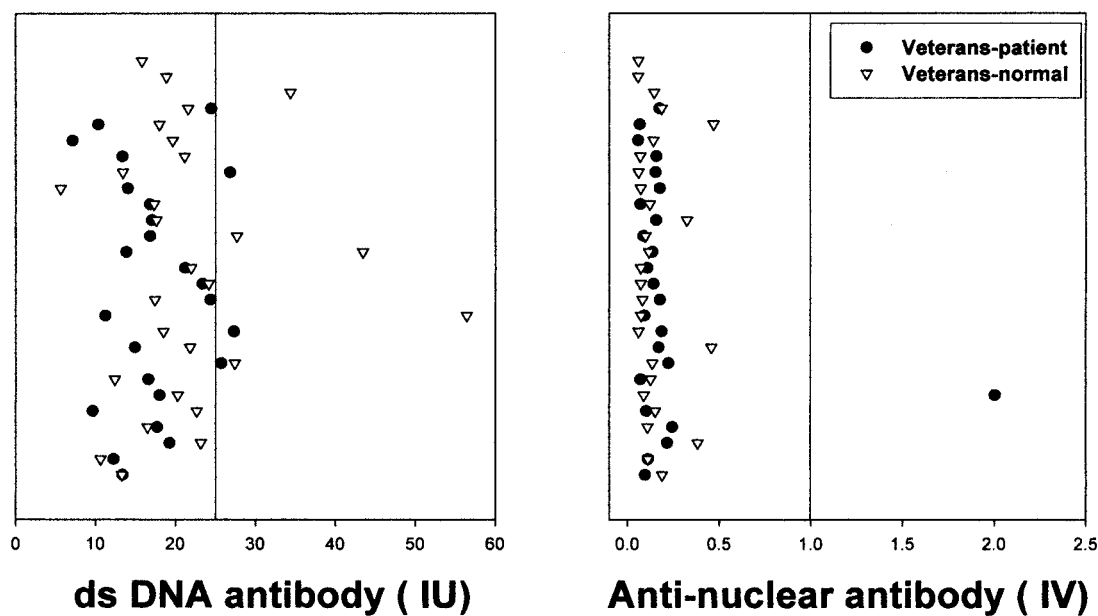


Fig. 1. No strong positive antibody responses to ds DNA and extractable nuclear antigens in the study subjects
Vertical line indicates the cut-off value for positive titer of anti-ds DNA (25 IU) or anti-ENA (1 IV).

antibody involved with induction of respiratory or dermal hypersensitive reactions^{25, 26}), were also enhanced in the veterans exposed to Agent Orange (Table 2), reflecting an atopic status. Unfortunately there was no previous report regarding effect of TCDD exposure on human plasma IgE levels, therein we could not conclude the IgE increase as a novel phenomena of TCDD exposure at this moment. However, one *in vitro* study addressing TCDD-induced enhancement of IgE synthesis from IL-4 plus CD40 mAb-stimulated human B cells²⁷) is supporting the result of our investigation. The involvement of IgE increase in the TCDD-mediated immune pathogenesis should be tested furthermore.

No obvious alteration was shown on composition and quantity of peripheral leukocyte population in the Vietnam War Korean veterans compared with that of the control subjects. Other studies, evaluating a relationship between changes in immunologic parameters and TCDD levels long after the end of exposure, did not find a significant effect of TCDD exposure on quantitative alteration of lymphocyte population including CD4⁺ T cell, CD8⁺ T cell, B lymphocyte, and natural killer cell^{11, 28, 29}). Regarding RBC parameters, there were decreases in the number of RBC, hemoglobin concentration, and hematocrit value for the veterans in comparison with those of the control subjects (Table 1). These findings suggest that RBC could be a target of TCDD-mediated immunotoxicity. But these decreases are not likely to be of clinical significance since the values are all within

the normal range. Even though very few data available for the hematopoietic effects of TCDD exposure in humans, one study described no difference in the RBC parameters listed above between the Vietnam War American veterans and their control³⁰). It will be difficult at the present moment to conclude whether TCDD exert its regulatory effects on RBC. It may be necessary to consider RBC as a potential target of TCDD-mediated toxicity and to further investigate TCDD effect on RBC hematopoiesis or clinical sequelae relevant to the present findings by recruiting much more study subjects in future.

The present study suggests that TCDD exposure may suppress host defense against infectious microorganisms, because IFN γ , a major cytokine mediating cell-mediated immune responses for the clearance of infectious pathogens, was significantly lowered in the veterans-patient group (Table 3). IFN γ is mainly produced by natural killer cells and type-1 helper and cytotoxic T cells, and induces macrophage activation leading to enhanced anti-microbial activity with IL-12^{31, 32}). Considering our *in vitro* experimental scheme using polyclonal T cell activator, activities of type-1 T cells may not be completely suppressed in the veterans exposed to TCDD, in that production of TNF α , another type-1-cytokine^{33, 34}), was not lowered in the veterans in comparison with that of the control group. In contrast to the IFN γ downregulation, production of type-2 cytokine, IL-4 and IL-10, was rather enhanced in the veterans (Table 3). Overall,

our results suggest that exposure to Agent Orange disturbs the homeostasis of T cell subsets resulting in predominance of type-2 response mediating by Th2 or Tc2 lymphocytes and concomitant suppression of type-1 T cell activities such as IFN γ synthesis. Combined with the our result on plasma IgE levels, our study indicates that immune system of the veterans-patient group is clearly skewing toward type-2 response. Orchestration of type-2 cytokines with IgE antibody could enhance the susceptibility of the veterans to various allergic diseases³⁵. Meanwhile, environmental toxicants such as lead, mercury, or diesel exhaust particles are known to preferentially drive the immune system toward type-2 response^{5, 18, 19, 36}. Exposure to these toxic chemicals may either contribute the upregulation of type-2 response in the veterans, or confound a substantial effect of TCDD on the balance between type-1 and type-2 response. Psychosocial stress could be considered a confounding factor contributing to the immune-modulation demonstrated at the Agent Orange-exposed veterans in our study. Changes in immune responses have been reported from peoples experiencing psychiatric problems through long-term patient care, examination, marital discord, or individual personal characteristics³⁷. An increased IL-10:IFN γ ratio from polyclonally stimulated PBMC was reported in the students stressed with examination³⁸, suggesting a potential association of stress hormones with shift toward type-2 response³⁹. Decrease in IgG antibody production after viral or bacterial vaccination was demonstrated to long-term caregivers of dementia patients^{40, 41}. Therein psychological stress rather than Agent Orange exposure may have a relationship with the immune-dysregulation of the veterans, resulting in skewing toward type-2 reactivity and lowered plasma IgG level. This explanation may be supported by recent results of the Korean Governmental epidemiologic study¹ addressing an association of mental depression or anxiety with self-rating level of Agent Orange exposure.

In conclusion, on the basis of our data with limitation on number of study subjects or information on exposure degree of Agent Orange, immune-alteration is apparently occurred in the Vietnam War Korean veterans. Even though we could not announce TCDD in the Agent Orange as a decisive factor for the disturbance of immune-homeostasis demonstrated in our study, military service in Vietnam and/or Agent Orange exposure could contribute the immune-dysregulation inducing the aberrant generation of immunoglobulins or cytokines. Dysregulation between type-1 and type-2 T cells shown in our study needs further extensive investigation to substantiate effect of TCDD exposure on cellular or humoral immunity in human.

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